Report

Origin and Diffusion of mtDNA Haplogroup X

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A maximum parsimony tree of 21 complete mitochondrial DNA (mtDNA) sequences belonging to haplogroup X and the survey of the haplogroup-associated polymorphisms in 13,589 mtDNAs from Eurasia and Africa revealed that haplogroup X is subdivided into two major branches, here defined as "X1" and "X2." The first is restricted to the populations of North and East Africa and the Near East, whereas X2 encompasses all X mtDNAs from Europe, western and Central Asia, Siberia, and the great majority of the Near East, as well as some North African samples. Subhaplogroup X1 diversity indicates an early coalescence time, whereas X2 has apparently undergone a more recent population expansion in Eurasia, most likely around or after the last glacial maximum. It is notable that X2 includes the two complete Native American X sequences that constitute the distinctive X2a clade, a clade that lacks close relatives in the entire Old World, including Siberia. The position of X2a in the phylogenetic tree suggests an early split from the other X2 clades, likely at the very beginning of their expansion and spread from the Near East.

mtDNA and the nonrecombining part of the Y chromosome are widely used in archaeogenetic studies (Renfrew 2000; Cavalli-Sforza and Feldman 2003) that aim

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1178

to reveal the human past. The uniparental inheritance and complete linkage of mutations in these two loci allow an unambiguous determination of the phylogenetic relationships between individual lineages. However, the putative genetic histories of the lineages that are obtained do not fully reflect the complex dynamics of ancient populations; thus, the data must be interpreted carefully. Phylogenetic clustering of mtDNA haplogroups has been found to be congruent with geography—there are haplogroups specific to African (Chen et

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Figure 1 Maximum parsimony phylogenetic tree of complete mtDNA sequences belonging to haplogroup X. Five mtDNAs were selected for complete sequence analysis in the course of the present study (Dr09, Es39, Gm66, Go41, and Om1538), 11 coding region sequences were from the work of Herrnstadt et al. (2002), but their control region sequences have now been added, and 5 complete sequences (Lev2, MM, E18, Nav125, and Oj2) were taken from the literature (Levin et al.1999; Maca-Meyer et al.2001; Mishmar et al. 2003; Bandelt et al., in press). The diagnostic mutations relative to the revised reference sequence (Andrews et al. 1999) are indicated on the branches. Transversions are specified by suffixes, and underlined mutations appear more than once in the tree. For protein-coding genes, "ns" indicates nonsynonymous and "s" synonymous mutations. For coalescence-time estimates, a mutation rate of 1 mutation per 5,139 years in the coding region (nps 577– 16023) was used (Mishmar et al. 2003). mtDNA sequence data are available at the Estonian Biocentre Web page.

al. 1995; Watson et al. 1997), Asian (Ballinger et al. 1992; Torroni et al. 1994; Kivisild et al. 2002), European/West Eurasian (Torroni et al. 1996; Macaulay et al. 1999), and Native American (Torroni et al. 1993) populations.

Haplogroup X is an exception to this pattern of limited geographical distribution. It is found, generally at low frequencies, in both West Eurasians (Richards et al. 2000) and some northern groups of Native Americans (Ward et al. 1991; Forster et al. 1996; Scozzari et al. 1997; Brown et al. 1998; Smith et al. 1999; Malhi et al. 2001), but, intriguingly, it is absent in modern north Siberian and East Asian populations (Brown et al. 1998; Starikovskaya et al. 1998; Schurr et al. 1999), which are genetically and geographically closest to those of Native

Americans. Among Siberians, haplogroup X mtDNAs have only been detected in some Altaian populations of southwestern Siberia (Derenko et al. 2001).

When the sequence variation of the first hypervariable segment (HVS-I) of the control region is analyzed, haplogroup X mtDNAs from Europe and the Near East are found to yield similar coalescence times: 17,000–30,000 years before present (YBP) and 13,700–26,600 YBP, respectively (Richards et al. 2000). These estimates are consistent with a pre-Holocene origin and spread of this haplogroup into West Eurasia. For Native Americans, the relatively old presence of haplogroup X is confirmed by the analysis of ancient human remains (Stone and Stoneking 1999; Malhi and Smith 2002). Moreover, Native American haplogroup X mtDNAs form a clade dis-

tinct from that of West Eurasians and with coalescence time estimates varying widely depending on both the method of estimation and the number of assumed founders. Thus, the coalescence times ranged from 12,000– 17,000 YBP to 23,000–36,000 YBP, times that are consistent with both a pre- and a postglacial population diffusion (Brown et al. 1998).

To obtain further information about the extent of haplogroup X diversity, 5 mtDNAs (from 1 Druze, 1 Estonian, 2 Georgians, and 1 Omani) were completely sequenced and were compared with 16 previously published X sequences (fig. 1). These latter sequences included the 11 haplogroup X coding sequences published by Herrnstadt et al. (2002) that have now been completed with the sequencing of the control region.

A maximum parsimony tree of the 21 haplogroup X sequences revealed that one nonsynonymous (13966) and three synonymous (6221, 6371, and 14470) substitutions in the coding region, as well as three transitions in the control region (153, 16189, and 16278), distinguish haplogroup X from the root of superhaplogroup N. Moreover, haplogroup X is subdivided into two major subhaplogroups, designated "X1" and "X2." Subhaplogroup X1, represented by a single Druze mtDNA in figure 1, differs from the root of haplogroup X by eight coding and three control region transitions and lacks the two transitions (195 and 1719) that characterize X2. These two nucleotides are rather mutable (Finnila¨ et al. 2001; Herrnstadt et al. 2002); thus, it cannot be completely ruled out that X1 is indeed a subset of X2 that reverted at both nucleotide positions. However, this possibility appears very unlikely, especially when one considers the time depth and the distinct geographic distribution of X1 (see below).

In contrast with X1, X2 is well represented in the tree, and it is further subdivided into at least six major clades (X2a–X2f), which include clades X1 and X2 as defined by Herrnstadt et al. (2002). All 20 X2 sequences, including the 2 Native American X2a sequences, share

transitions at nucleotide positions (nps) 195 and 1719. A recurrence of the nonsynonymous substitution at 13708 was observed (clades X2b and X2d). In addition, the nonsynonymous transition 13966 was found to have reverted in the Moroccan X2b sequence. These were the only recurrent mutations found among the 67 variable positions in the coding region sequences. The ratio of nonsynonymous to synonymous substitutions was 0.40 (18/45). Six mutations were located in RNA genes.

The data obtained from the analyses of complete mtDNA sequences belonging to haplogroup X were then used to survey 13,589 mtDNAs (21,682 when mtDNA data from the literature are included) from 66 populations of Eurasia and North Africa (table 1). A total of 175 mtDNAs were found to harbor the four coding region transitions—at 6221, 6371, 13966, and 14470 that define haplogroup X. The four markers were always found in association and, by combination of the control and coding variation, all 175 mtDNAs could be apportioned to subhaplogroups X1 and X2 (table 2). No other main branches occur, and the root haplotype was not present among the sequences. The adenine at np 153 appears relatively conserved in the phylogenetic context of 376 complete human mtDNA sequences taken worldwide, exhibiting a change to guanine only in haplogroups A and X (Ingman et al. 2000; Finnilä et al. 2001; Maca-Meyer et al. 2001; Derbeneva et al. 2002; Mishmar et al. 2003). In contrast, this position shows a high level of variation in the background of haplogroup X as both the 153A and 153G alleles are present in its different subclades. It is possible that the $A\rightarrow G$ transition at 153 arose only once in the haplogroup X ancestor and the recurrent reverse mutations in 11 branches in figure 2 bear witness to the process of favored fixation of the more stable A allele.

Subhaplogroup X1 was found to be largely restricted to the Afro-Asiatic–speaking populations of North Africa and neighboring areas, including Ethiopia, suggesting a possible geographic diffusion of X1 alongside the

Figure 2 Median-joining (MJ) network of 175 haplogroup X partial sequences. The MJ (Bandelt et al. 1999) algorithm was implemented within the Network 3111 program (A. Röhl; Shareware Phylogenetic Software Web site). Default settings were used for HVS-I (nps 16024– 16383), HVS-II (nps 16518–310), and coding-region sequence variation as given in table 2. Highly variable (Hasegawa et al. 1993) positions of HVS-I (16093, 16126, 16129, 16187, 16189, 16223, 16234, 16278, 16292, 16293, 16311, 16325, 16355, and 16362) were assigned a weight of 1, other HVS-I and HVS-II sites were assigned a weight of 2, and coding-region sites were assigned a weight of 10. Areas of the circles are proportional to haplotype frequencies. Populations are indicated by the following abbreviations: ab = Abazin, ad = Adygei, alb = Albanian, alg = Algerian, alt = Altaian, ar = Armenian, arb = Arab from Uzbekistan, be = Bengali, cz = Czech, dr = Druze, eg = Egyptian, es = Estonian, et = Ethiopian, ev = Evenk, fr = French, go = Georgian, gr = Greek, ho = Croat, hu = Hungarian, ir = Iranian, it = Italian, jo = Jordanian, kir = Kyrgyz, kr = Karachay, ku = Kumyk, kw = Kuwaiti, le = Lebanese, mo = Moroccan, ng = Nogay, $os =$ Ossetian, sa = Saudi Arabian, sw = Swede, sy = Syrian, td = Tadjik, tu = Turk, and ukr = Ukrainian. Variant bases are numbered (Anderson et al. 1981) and are shown along links between haplotypes. Nucleotide changes are specified by suffixes only for transversions, and a "d" indicates a deletion. The node marked with a large asterisk (\bigstar) matches the root type of haplogroup X. The coalescence times of the clades are shown near the clades. Coalescence times of HVS-I clusters were calculated by means of ρ , the average mutational distance to the founder haplotype of the cluster, by using a mutation rate of 1 transition per 20,180 years in the segment between nps 16090 and 16365 (Forster et al. 1996). Standard deviations for ρ were calculated as in the work of Saillard et al. (2000), a procedure which ignores the variance due to molecular clock calibration. mtDNA sequence data are available at the Estonian Biocentre Web page.

Table 1

(*continued*)

Table 1 (continued)

^a An asterisk (*) denotes frequencies deduced from published HVS-I sequences. Populations are divided into regions basically as in (Richards et al. 2000), with some changes: North African and South Caucasian populations were considered separately from populations of the Near East, and all European populations of the Mediterranean area were aggregated into a single Mediterranean European region. Sources are denoted as follows: 'present study, 'Bermisheva et al. (2002), 'Bertranpetit et al. (1995), 'Corte-Real et al. (1996), ^sCrespillo et al. (2000), ⁶Derbeneva et al. (2002), ⁷Derenko et al. (2001), ⁸Dimo-Simonin et al. (2000), ⁹Di Rienzo and Wilson (1991), ¹⁰Finnilä et al. (2001), ¹¹Helgason et al. (2000), ¹²Helgason et al. (2001), ¹³Hofmann et al. (1997), ¹⁴Kivisild et al. (1999), ¹⁵Kivisild et al. (2003), 16 Lutz et al. (1998), 17 Macaulay et al. (1999), 18 Malyarchuk et al. (2002), 19 Mogentale-Profizi et al. (2001), 20 Opdal et al. (1998), 21 Orekhov et al. (1999), 22Parson et al. (1998), 23Pereira et al. (2000), 24Pereira et al. (2001), 25Pfeiffer et al. (1999), 26Piercy et al. (1993), 27Rando et al. (1998), 28Richards et al. (1996), 29Richards et al. (2000), 30Rousselet and Mangin (1998), 31Saillard et al. (2000), 32Salas et al. (2002), 33Tagliabracci et al. (2001).

^b The 95% credible regions (CR), calculated using software kindly provided by Vincent Macaulay, are shown in brackets.

 c Haplotype diversities were calculated as in the work of Nei (1987), only for regions where more than six haplogroup X mtDNAs were available considering the sequence variation observed between nps 16090 and 16365.

Table 2

 $(continued)$

Table 2 (continued)

Table 2 (continued)

^a Two hypervariable segments of the control region were sequenced: HVS-I (16024-16383) and HVS-II (16518-310). Nucleotide change is specified for transversions; d = deletion. Length variation in the C stretch (16184-16193) is not shown. Mutations that were sequenced outside the specified range are shown in parentheses. Sequence analyses were performed using the Sanger dideoxy chain-termination method with the Amersham DYEnamic ET Terminator Cycle Sequencing Kit (Amersham Pharmacia Biotech) on an ABI PRISM 377 DNA Sequencer. Sequences were alignedand analyzed with the Wisconsin Package (GCG).

Restriction-endonuclease sites are indicated as follows: c = Ddel, e = HaelII, g = HinfI, i = MspI, j = MboI, k = RsaI, l = TaqI, m = BamHI, s = AccI,t = BstOI, u = MseI, w $= MboII$, $y = Bfml$. Haplogroup assignments are according to figure 2.

Populations are indicated by the following letter code: ab = Abazin, ad = Adygei, alb = Albanian, alg = Algerian, alt = Altaian, ar = Armenian, arb = Arab from Uzbekistan, be = Bengali, $cz = Czech$, dr = Druze, eg = Egyptian, es = Estonian, et = Ethiopian, ev = Evenk, fr = French, gm and go = Georgian, gr = Greek, ho = Croat, hu = Hungarian, ir = Iranian, it = Italian, jo = Jordanian, kir = Kyrgyz, kr = Karachay, ku = Kumyk, kw = Kuwaiti, le = Lebanese, mo = Moroccan, ng = Nogay, os = Ossetian, sa = Saudi Arabian, $sw =$ Swede, sy = Syrian, td = Tadjik, tu = Turk, ukr = Ukrainian.

HVS-I nucleotide positions are -16000 .

^e All listed HVS-II sequences also harbored the transitions 16519, 73, and 263.

^f Revised Cambridge reference sequence (Andrews et al. 1999).

Mediterranean Sea and the Red Sea (table 1). This subhaplogroup is subdivided into the two clades X1a and X1b, which are defined by two and five coding region mutations, respectively (fig. 2). Both clades share a recurrent transition at 146 in HVS-II. The coalescence time of the entire X1 subhaplogroup using HVS-I variation is 42,900 \pm 18,100 YBP, whereas the coalescence time of the X1a clade is $17,900 \pm 11,900$ YBP.

Virtually all (97.2%) haplogroup X mtDNAs from the Near East, the South Caucasus, and Europe were found to belong to subhaplogroup X2, as did all (100%) of those from Siberia and Central Asia and some (36.8%)

of those from North Africa (table 2). Thus, subhaplogroup X2 is characterized by a very wide geographic range but also by an infrequent occurrence. Indeed, it generally comprises $\langle 5\%$ of the mtDNAs in West Eurasian and North African populations (table 1). Three exceptions include the Druze, the Georgians, and the Orkney Islanders, among whom the frequency of X2 reaches 11%, 8%, and 7%, respectively. The high frequencies of X2 in the Druze and the Orkney Islanders are combined with a low haplotype diversity (0.400 and 0.473, respectively), and the relatively high frequency in these populations is most likely due to genetic drift and founder events. Overall, it appears that the populations of the Near East, the Caucasus, and Mediterranean Europe harbor subhaplogroup X2 at higher frequencies than those of northern and northeastern Europe (P < .05) and that X2 is rare in Eastern European as well as Central Asian, Siberian, and Indian populations and is virtually absent in the Finno-Ugric and Turkic-speaking people of the Volga-Ural region. Coalescence time estimates based on HVS-I and coding region variation— $17,900 \pm 2,900$ YBP and $21,600 \pm 4,000$ YBP, respectively (figs. 1 and 2)—are consistent with the range expansion of X2 around or after the last glacial maximum (LGM). It is intriguing that the estimated coalescence time for X2 alone is very close to that obtained from HVS-I data for the entire haplogroup X (20,200 \pm 3,100 YBP) (fig. 2). However, the latter is probably an underestimate due to both the higher proportion $(>90\%)$ of X2 mtDNAs included in the calculations and the fact that the HVS-I consensus sequence of X2 is completely identical to that of the overall haplogroup X.

Two-thirds of the subhaplogroup X2 sequences fall into the five clades X2b–X2f (fig. 2). Two sequences one from Lebanon and one from Georgia—lacked the transition at np 1719, suggesting either the presence of an early X2 branch or a reversion at that nucleotide position. The sister groups X2b and X2c (X1 and X2, respectively, in the work of Herrnstadt et al. 2002) encompass one-third of the European sequences (excluding the samples from the North Caucasus). It is of interest that some North African sequences (from Morocco and Algeria) belong to X2b as well. Subhaplogroup X2b shows a diversity that is consistent with a postglacial population expansion in both West Eurasia and North Africa. Clades X2e and X2f encompass the majority (87.1%) of the sequences from the South Caucasus area and show coalescence times (12,000 \pm 4,000 YBP and $10,800 \pm 5,000$ YBP, respectively) consistent with a Late Upper Paleolithic (LUP) origin and a subsequent spread in the region. We found significant differences between the haplogroup distribution between the North and the South Caucasian samples, a result that indicates a major geographical barrier between the two regions. The South Caucasian sample is enriched in mtDNAs belonging to clades $X2e$ and $X2f$ ($P < .01$), whereas the North Caucasian sample shows a higher proportion of sequences derived at nps 225 and 16248 $(P < .01)$.

Clade X2e, defined by the synonymous substitution at 15310, encompasses all haplogroup X sequences in the Altaians (fig. 2). Among the nine Altaian X sequences, eight harbor the founder HVS-I motif of X2e, and seven of these eight also carry the HVS-II founder motif. As a result, a very low haplotype diversity of haplogroup X (0) in the Altaian region was obtained, making it significantly different from the haplotype di-

versities for haplogroups C and D (0.835 and 0.943, respectively) in the same region. Moreover, the nine Altaian mtDNAs do not harbor any nucleotide difference between nps 16090 and 16365. Therefore, under the assumption that these sequences are a random sample of the Altaian haplogroup X, an estimated ρ value <0.33 $(P < .05)$ was obtained. This value corresponds to a time depth of $\lt6,700$ years (Forster et al. 1996), and it would suggest that Altaians have acquired haplogroup X2 only relatively recently. This scenario is supported by the concomitant presence in the Altaians of a wide range of other West Eurasian haplogroups (H, J, I, T, U1, U4, and U5) that comprise ∼27% of their mtDNAs. Indeed, any recent migration (for example, from the [southern] Caucasus region) that might have carried X2e mtDNA sequences to the Altai region would also explain the presence of the other West Eurasian mtDNA haplogroups in modern Altaians.

Further northeast of the Altai area, haplogroup X sequences were detected in the Tungusic-speaking Evenks, of the Podkamennaya Tunguska basin (Central Siberia). In contrast to the Altaians, the Evenks did not harbor any West Eurasian mtDNA haplogroups other than X. However, neither of the two Evenk X haplotypes showed mutations characteristic of the Native American clade X2a. Instead, one sequence was a member of X2b and the other of $X2^*$ (fig. 2). Thus, one possible scenario is that several X haplotypes arrived in Siberia from western Asia during the Palaeolithic, but only X2a crossed Beringia and survived in modern Native Americans. Alternatively, the presence of two phylogenetically different haplogroup X mtDNA sequences in this particular subpopulation of Evenks might be due to recent gene flow.

The Native American–specific clade X2a appears to be defined by five mutations, three in the coding region (8913, 12397, and 14502) and two in the control region $(200 \text{ and } 16213)$ (fig. 1). The transition at np 200 was seen in virtually all previously analyzed Native American haplogroup X mtDNAs, whereas the transition at np 16213 was absent in some of the Ojibwa described by Brown et al. (1998). We surveyed our Old World haplogroup X mtDNAs for the five diagnostic X2a mutations (table 2) and found a match only for the transition at np 12397 in a single X2* sequence from Iran. In a parsimony tree, this Iranian mtDNA would share a common ancestor with the Native American clade (fig. 2). Yet, the nonsynonymous substitution at np 12397 converting threonine to alanine cannot be regarded a conservative marker, as it has also been observed in two different phylogenetic contexts—in haplogroups J1 and L3e—among 794 complete mtDNA sequences (Finnilä et al. 2001; Maca-Meyer et al. 2001; Herrnstadt et al. 2002). Therefore, the scenario that the threonine to alanine change in the haplogroup X background is indeed due to recurrence appears most plausible.

These findings leave unanswered the question of the geographic source of Native American X2a in the Old World, although our analysis provides new clues about the time of the arrival of haplogroup X in the Americas. Indeed, if we assume that the two complete Native American X sequences (from one Navajo and one Ojibwa) began to diverge while their common ancestor was already in the Americas, we obtain a coalescence time of 18,000 \pm 6,800 YBP, implying an arrival time not later than 11,000 YBP.

The results of this study point to the following conclusions. First, haplogroup X variation is completely captured by two ancient clades that display distinctive phylogeographic patterns—X1 is largely restricted to North and East Africa, whereas X2 is spread widely throughout West Eurasia. Second, it is apparent that the Native American haplogroup X mtDNAs derive from X2 by a unique combination of five mutations. Third, the few Altaian (Derenko et al. 2001) and Siberian haplogroup X lineages are not related to the Native American cluster, and they are more likely explained by recent gene flow from Europe or from West Asia. Fourth, the split between "African" X1 and "Eurasian" X2 subhaplogroups of X is phylogenetically as deep as that within the branches of haplogroup U that also differ profoundly in their phylogeography. Thus, subhaplogroup U6 is largely restricted to North Africa (as X1), whereas subhaplogroup U5 is widespread in West Eurasia (as X2). The phylogeographic patterns and the coalescence times that we obtained here suggest that the basic phylogenetic structures of the mtDNA haplogroups in West Eurasia and North Africa are as ancient as the beginning of the spread of anatomically modern humans in this region. Finally, phylogeography of the subclades of haplogroup X suggests that the Near East is the likely geographical source for the spread of subhaplogroup X2, and the associated population dispersal occurred around, or after, the LGM when the climate ameliorated. The presence of a daughter clade in northern Native Americans testifies to the range of this population expansion.

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Electronic-Database Information

Accession numbers and URLs for data presented herein are as follows:

- Estonian Biocentre, http://www.ebc.ee/EVOLUTSIOON/ mtDNA-X/ (for mtDNA sequence data)
- Shareware Phylogenetic Network Software, http://www .fluxus-engineering.com/sharenet.htm (for Network 3111)

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